

Further studies of the reactivity of chlorocarbene and the different behavior of methylene bromide toward butyllithium will be the subject of a detailed publication.

DEPARTMENT OF CHEMISTRY
THE UNIVERSITY OF CHICAGO
CHICAGO 37, ILL.

GERHARD L. CLOSS
LISELOTTE E. CLOSS

RECEIVED JULY 9, 1959

CHROMATOGRAPHY OF MYOSIN

Sir:

The general method of Peterson and Sober¹ has been applied to the muscle protein, myosin or "myosin A." Myosin A^{2,3,4} freed of myosin B by dialysis against 0.2 M KCl, 0.01 M tris pH 7.4 in the presence of adenosine triphosphate and by 1 hour of centrifugation at 55,000 × g was passed through a diethylaminoethyl cellulose column equilibrated with a solvent 0.2 M KCl, 0.01 M tris pH 7.4. An ascending gradient to 1.0 M KCl was applied (Fig. 1), and protein concentration was measured⁵ in the effluent. Protein recovery was better than 80%.

TABLE I

Prepn.		α	β
19	$\bar{M}_w \times 10^{-5}$	4.52	6.10
	\bar{r}_g	437	474
	V_m (2 d.)	4.7	9.5
22	$\bar{M}_w \times 10^{-5}$	4.55	5.00
	\bar{r}_g	434	560
	V_m (12 d.)	0.4	3.8
28	$\bar{M}_w \times 10^{-5}$	4.02	5.60
	\bar{r}_g	475	634
	V_m (3 d.)	5.0	17
33	$\bar{M}_w \times 10^{-5}$	4.21	6.36
	\bar{r}_g	430	500
	V_m (0 d.)	8.0	8.7
21	$\bar{M}_w \times 10^{-5}$	4.00	..
	\bar{r}_g	434	..

Myosin is resolved into at least two components, α and β (Fig. 1). Neither component shows a turbidity drop on adenosine triphosphate addition, confirming the elimination of myosin B. The α -component probably is highly purified myosin. The data⁶ of Table I yield an average \bar{M}_w of 4.3×10^5 g. and an average \bar{r}_g of 442 Å. \bar{M}_w from ultracentrifuge work⁷ is 4.2×10^5 g. This shows that the two methods can agree; moreover the straightness of the Zimm light-scattering plot (Fig. 1) does not encourage speculation about myosin non-uniform substructure.⁸ In this work the "full" Zimm plot (*i.e.*, intensities at various concentra-

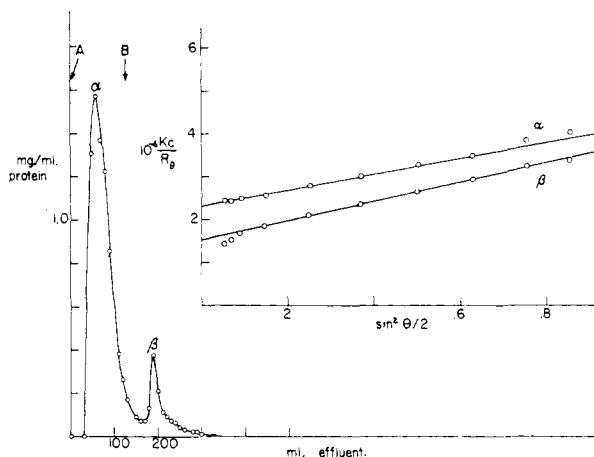


Fig. 1.—Chromatography on a 13 × 2.5 cm. column of diethylaminoethyl cellulose (1 meq./g.); eluting solutions: A—0.2 M KCl, 0.01 M tris pH 7.4; B—gradient elution to 1.0 M KCl; flow rate 60 ml./hr.; 10 ml. fractions were collected. The gradient used was composed of two conical vessels filled with 250 ml. of 1.0 M KCl, 0.01 M tris pH 7.4 and 125 ml. of 0.2 M KCl, 0.01 M tris pH 7.4. Insert shows: Zimm plot of α and β fractions in 0.5 M KCl, 0.01 M tris pH 7.4.

tions as well as at various angles) was not attempted because it has been shown⁸ that in 0.6 M KCl the second virial coefficient is essentially zero. The β -component is heavier (average \bar{M}_w , 5.77×10^5 g.) and more extended (average \bar{r}_g , 542 Å.); also its specific ATPase activity,⁹ V_m , (Table I) is greater and more thermostable than that of the α -component. Scattered observations suggest that β may be transformable into α , either by warming briefly from 4 to 25°, or by aging.

The author is indebted to Dr. M. Gellert for guidance in light-scattering measurements, to Dr. M. F. Morales for general counsel, and to Dr. W. Niemierko for valuable criticisms. This work was supported by a Rockefeller Fellowship and by Training Grant 2G-174 of the U.S.P.H.S.

(9) μ mole P-sec.⁻¹ g. protein⁻¹ in 0.5 M KCl, 0.1 M tris, 10^{-3} CaCl₂, pH 8.0, 25°. The age of myosin preparation (in days) is indicated in parentheses.

DEPARTMENT OF BIOCHEMISTRY
DARTMOUTH MEDICAL SCHOOL
HANOVER, N. H.

J. BRAHMS

RECEIVED JULY 6, 1959

OPTICAL ROTATORY DISPERSION STUDIES. XXX.¹ DEMONSTRATION OF BOAT FORM IN A 3-KETO STEROID²

Sir:

Kinetically controlled bromination of 2 α -methylcholestan-3-one³ (or of its enol acetate) leads to 2-bromo-2-methylcholestan-3-one (m.p. 136–138°), whose spectral properties ($\lambda_{\max}^{\text{CHCl}_3}$ 5.84 μ ; $\lambda_{\max}^{\text{cyclohex}}$ 313 m μ) require⁴ an axial bromine atom. By

(1) Paper XXIX, P. Crabbé, C. Djerassi, E. J. Eisenbraun and S. Liu, *Proc. Chem. Soc.*, in press.

(2) Supported by grant No. CY-2919 from the National Cancer Institute.

(3) Y. Mazur and F. Sondheimer, *THIS JOURNAL*, **80**, 5220 (1958).

(4) (a) R. N. Jones, D. A. Ramsay, F. Herling and K. Dobriner, *ibid.*, **74**, 2828 (1952); (b) R. C. Cookson, *J. Chem. Soc.*, 282 (1954).

(1) E. Peterson and H. A. Sober, *THIS JOURNAL*, **78**, 751 (1956).

(2) A. Szent-Györgyi, "Muscular Contraction," Academic Press, Inc., New York, N. Y., 1947.

(3) Dr. J. Botts, private communication.

(4) H. H. Weber and H. Portzehl, *Advances in Protein Chemistry*, **7**, 161 (1952).

(5) O. H. Lowry, N. J. Rosebrough, A. L. Farr and R. J. Randall, *J. Biol. Chem.*, **133**, 265 (1951).

(6) For specific refractive index increment the value of 0.209 ml./g. was used.

(7) P. H. von Hippel, H. K. Schachman, P. Appel and M. F. Morales, *Biochim. Biophys. Acta*, **28**, 504 (1958).

(8) A. Holtzer and S. Lowey, *THIS JOURNAL*, **81**, 1370 (1959).